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Short communication

Use of porous pyrolytic carbon for analytical and microscale highperformance liquid chromatographic bioseparations

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Abstract

Porous carbonaceous adsorbent was prepared by carbonization of saccharose in silica gel pores followed by leeching out of the silica matrix. The product of pyrolysis was then deactivated by hydrogenation. The resulting adsorbent shows intermediate sorption properties between those of the highly polar pyrolytic glassy carbon and the hydrophobic graphitized carbon. The microparticulate mesoporous carbon was examined for its use in capillary HPLC separations. The separation of selected stereoisomers in a 320 μ m I.D. capillary column packed with the porous carbon particles is described and discussed. Additionaly, the porous carbon filled with dextran gel was tested as a material for direct HPLC analysis of drugs in human serum. \circ 1998 Elsevier Science B.V. All rights reserved.

Keywords: Bioseparations; Porous pyrolytic carbon

porous carbon has been growing during the last show high chemical stability over a wide pH range, decade. The most commonly used methods of prepa- adequate mechanical stability to withstand abrasion ration of porous carbon particles that are suitable for and high pressures, as well as sufficient surface area, HPLC are the controlled carbonization of the poly- high pore volume and controlled pore size distribumeric precursor particles at high temperatures [1,2] tion [5–7]. and the method in which the pores of appropriate Carbonization of the organic precursors below porous matrix are filled with a carbon precursor 1000°C results in an amorphous pyrolytic carbon which is subsequently pyrolysed [3,4]. The auxiliary containing micropores and mesopores. It possesses

1. Introduction materials such as the starting porous sorbent or porogenes are leeched out after the carbonization of The acceptance of separation media based on the precursor. The products of carbonization usually

oxygenated surface bearing various functional groups *Corresponding author.
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pyrolytic carbon is heated to about 1500°C, amorway, Santa Clara, CA 95051, USA. phous glassy carbon is produced. In turn, carbon

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heated above 2000°C has the atomic structure of largest contribution to adsorption on the carbonacetwo-dimensional graphite. Each of the above men- ous surface seems to be associated with the dispertioned materials shows different adsorption prop- sion forces of the sorbate molecules. erties. Although the carbon packing materials provide

tures acts as a very strong polar adsorbent. However, column efficiency for carbon-packed analytical col-
a high sorption activity of the pyrolytic carbon often umns usually does not match that obtained for well a high sorption activity of the pyrolytic carbon often causes undesired irreversible retention [7]. The established silica-based materials. Therefore, a pyrolytic carbon has been successfully used for miniaturization of the column size, together with purification and preconcentration of analytical sam- improving the quality of the packing material can ples [8] and, after appropriate deactivation, also for increase the performance of carbon packed columns.
HPLC separations [9]. In contrast, the porous Microcolumns and capillaries are becoming popular HPLC separations [9]. In contrast, the porous Microcolumns and capillaries are becoming popular graphitic carbon prepared at very high temperatures in both electrochromatography [22] and capillary has a hydrophobic and chemically homogeneous HPLC [23]. surface. Its retention properties can be compared For example, polymer-coated open-tubular microwith those of commercial ODS-bonded silica phases columns [24,25] or capillaries packed with silica-[10–13]. based particles [26,27] provide fast and low-volume

the pyrolyzed carbon can be obtained not only via biological samples. Narrow-bore columns packed the high-temperature treatment at above 2000°C, but with a carbonaceous adsorbent were described previthe high-temperature treatment at above 2000° C, but also by a chemical modification of highly-active ously [28]; however, separations on a carbonaceous carbon surface. For example, hydrogenation of oxy- adsorbent in the microcolumns with an internal gen-containing functional groups at about $1000^{\circ}C$ [6] diameter smaller than 1 mm have not been reported or immobilization of a layer of inactive polymer on yet. the carbon surface [14] have been used for deactiva- In the present paper we describe capillary HPLC tion. \blacksquare the separations of stereoisomers using a 320 μ m I.D.

modification, in which a polysaccharide gel was croparticulate porous carbon. Additionaly, the created within the pores of carbon adsorbent [15]. pyrolytic carbon modified with crosslinked dextran The polysaccharide network prevents larger solutes has been tested for its use in direct analysis of drugs from contact with the carbon, while smaller mole- in biological fluids. cules may penetrate the network and interact with the original carbon surface.

Both high specific surface area and a special **2. Experimental** morphology of the surface make pyrolytic or graphitic carbon adsorbents sensitive not only to the 2.1. *Materials* chemical nature of chromatographed compounds but also to their size, shape and position of function- Mesoporous pyrolytic carbon (CF-35) was prealities. It is known that carbon-based adsorbents pared according to the silica template method [6]. show unique selectivity towards positional isomers Saccharose was carbonized at 650° C in the pores of [16,17], and *cis*–*trans* isomers [9,18–20]. A sepa- the silica gel matrix having an average pore diameter ration of enantiomers using a carbon column has of 6 nm. Silica was then dissolved in aqueous been illustrated [21]. The unique separation prop-
sodium hydroxide. Finally, the spheroidal carbonaceerties of carbonaceous adsorbents could be explained ous material with replicated porous structure was by a combination of various kinds of selective dried and deactivated by hydrogenation at 1000°C interactions between solutes, mobile phases and [6,7]. Dextran-shielded carbon adsorbent was prestationary phases such as electron donor–acceptor pared by filling the pores of original carbon CF-35 interactions, hydrogen bonding, etc. However, the with crosslinked dextran. The crosslinking of dextran

Carbon prepared by pyrolysis at lower tempera- unique selectivity for numerous separations, the in both electrochromatography [22] and capillary

A better uniformity of the surface composition of analyses of isomeric compounds or complicated

Recently, we introduced a new procedure of fused-silica capillary column packed with the mi-

was performed under alkaline conditions using **3. Results and discussion** butandiepoxide reagent as described elsewhere [15].

Samples of α -irone isomers were prepared at 3.1. Properties of carbonaceous adsorbent Faculty of Pharmacy, Turin, Italy, and ketoprofen was purchased from Sigma (St. Louis, MO, USA). HPLC packing material used in the study consists The other chemicals and solvents, purchased from of potato-shaped particles of pyrolytic carbon with various commercial sources, were of HPLC or an average size between 10 and 20 μ m. The pore analytical reagent grade and used as received. size varies from several nm up to several hundreds

on a system equipped with a Kontron 322 pump material. Large and very active surface bearing a (Kontron Instruments, Milan, Italy), an Acurate 70 variety of functionalities supposedly causes strong flow-rate converter (LC-Packings), and a Rheodyne retention of chromatographed compounds with dif-7125 injector (Rheodyne, Berkeley, CA, USA) pro- ferent chemical structures. In the system studied by vided with a 20 μ l polyether ether ketone (PEEK) us, the elution strength of a mobile phase for solutes loop. The flow-rate converter was set up at 1:70 of medium polarity increased in the sequence from (inlet:outlet flow) which gave a constant flow-rate of water, methanol, acetonitrile to tetrahydrofuran. This 14.5 ml/min. The whole volume of the loop was makes the situation similar to typical reversed-phase injected into the capillary column equilibrated with a chromatography. However, separation on the water-rich starting eluent. Then, the compounds pyrolytic carbon studied did not obey the theory of concentrated on the top of the column were eluted by conventional retention developed for reversed-phase a gradient of increasing concentration of acetonitrile chromatography, e.g. a logarithm of retention factor in the mobile phase. Detection was carried out with a was not a linear function of eluent composition. To Kontron 433 UV capillary detector, equipped with a completely elute some hydrophobic compounds, Z-shape flow-microcell (total volume of 90 nl, especially those with condensed aromatic rings, an optical path length of 20 mm) or alternatively with a extremely nonpolar eluent such as hexane had to be Kontron 440 DAD-UV unit equipped with a capillary used. On the other hand, highly polar compounds flow-cell (total volume of 200 nl). such as phenols, interacting strongly with polar

direct injection analysis of drugs in serum was dipole–dipole interactions, were retained by column performed with an HPLC system consisting of using nonpolar eluents and could be eluted from the Kontron 322 pump, Rheodyne 7160 injector, and column with more polar mobile phases, thus resem-Kontron 440 DAD-UV detector equipped with an bling the normal-phase HPLC. The mixed character analytical flow-cell. $\qquad \qquad$ of the carbon surface can cause peak tailing and

nm. The sponge-like structure of carbon matrix

2.2. *Packing the columns* esults in a high specific surface area of 940 m²/g

(nitrogen adsorption-BET), while its good mechani-A fused-silica capillary column Fusica (150 mm × cal strength is preserved. The physical and chemical
320 μ m I.D.) was custom-packed by LC-Packings
International, (Amsterdam, The Netherlands). Ana-
lytical stainless ste

amount of aromatic and oxygen-containing groups is 2.3. *LC experiments* situated on the pyrolythic carbon surface. Therefore, a mixed behaviour owing to the presence of both The capillary HPLC separations were performed polar and nonpolar sites is typical for such a Conventional analytical chromatography used for groups of carbon surface via hydrogen bonding or Data from both HPLC systems were acquired and complicate the optimization of separation. However, processed by a Kontron data system 450-MT2/DAD. in certain cases, it may help to discriminate the separate. trile–water (80:20, v/v). The sensitivity of the

carbon adsorbent shows high selectivity for sepa- cules. In the case of α -irone, the *E*-isomer molecule ration of *cis*–*trans* (*E*/*Z*) isomers [9]. Various *E*/*Z* is more planar and therefore provides a larger contact isomers have been separated using commercially area with the carbonaceous adsorbent. As a result, available or laboratory made columns based on the *E*-isomer is more strongly retained than the graphitized carbon [18–20]. We used our carbon- *Z*-isomer. packed capillary for the separation of *E*/*Z* isomers of an a-irone. An extract containing a-irone is used for 3.3. *Direct serum injection* preparation of perfumes. The separation may help to determine the quality of a perfume, since particular Proteins are irreversibly adsorbed on the pyrolytic isomers differ significantly in their scent. carbon surface. Therefore, protein-containing sam-

separated using the capillary-HPLC column packed directly, but only after an expensive and time with the micrometer-sized pyrolytic carbon. The consuming preseparation step. To prevent proteins separation with a selectivity of 6.63 was achieved from irreversible adsorption on a carbonaceous adunder simple isocratic conditions in about 15 min. sorbent it must be chemically deactivated, e.g. by a Since only a very diluted sample was available, 20 layer of hydrophilic polymer [14]. However, the ml of the sample was let to concentrate on the top of carbon coated with an inert layer often loses its

compounds which are otherwise very difficult to the capillary column and then eluted with acetonimethod can be increased significantly using on-line 3.2. *Separation of cis*–*trans isomers* sample preconcentration without sacrificing the selectvity of separation. The retention depends on the It has been previously found that the pyrolytic difference in the geometric structure of the mole-

Fig. 1 shows that the isomers can be easily ples such as serum cannot be chromatographed sorption activity and separation selectivity also for small analytes. In order to obtain a sorbent which would selectively adsorb only the compounds of interest from a multicomponent sample, we filled the pyrolytic carbon with a dextran gel of a specific pore size [15]. The modification allows small molecules to interact with the carbon surface after penetration the hydrophilic dextran layer, while large molecules are prevented from contact with the active surface because of their steric exclusion from the dextran gel pores. As a result, protein molecules should pass through the column unretained. The mechanism is typical for 'restricted access media' [29].

To test the dextran-modified carbon for its use in direct serum analysis, we determined recoveries of various proteins injected. Recoveries between about 80 and 100% were found for proteins of both acidic and alkaline nature, such as chymotrypsinogen A and ovalbumin, and of different molecular mass ranging from 14 500 (ribonuclease A) to 675 000 (thyro-Fig. 1. Capillary HPLC of *cis* and *trans* isomers of α -irone. globulin). Pure human serum, ketoprofen and the Conditions: column, fused-silica capillary, 150 mm \times 320 μ m I.D.; corum spiked with the various amount Conditions: column, fused-silica capillary, 150 mm \times 320 μ m 1.D.;
packing material, pyrolytic carbon CF-35; mobile phase, acetoni-
trile–water (80:20, v/v); flow-rate, 14.5 μ /min; detection, abwere injected repeate sorbance at 254 nm; injection, 20 μ l of sample dissolved in umn packed with the dextran-modified carbon to test acetonitrile–water (30:70, v/v). its ability to separate the drug from the serum. Fig. 2

Fig. 2. HPLC of ketoprofen in serum on dextran-shielded carbon rations of *cis–trans* isomers and enantiomers.

column. Conditions: column stainless steel, 100×4 mm I.D.; column. Conditions: column stainess steel, 100 λ 4 mm I.D.; Additionally, we showed that carbon with pores packing material, pyrolytic carbon CF-35 with pores filled with croslinked dextran can be used as an deveran call dextran gel; mobile phase, acetonitrile-water (10:90, v/v), followed by a step gradient to acetonitrile–water $(70:30, v/v)$, in the alternative material for direct HPLC analysis of third minute from injection; flow-rate, 1.0 ml/min; detection, drugs in serum. absorbance at 254 nm; sample, human serum spiked with $2 \mu g/ml$ of ketoprofen; injection volume, $20 \mu l$.

shows a typical chromatogram of the mixture. All proteins were eluted from the column using acetoni- [1] I. Novák, D. Berek, CS patent 230 (1984) 297. trile–water $(10:90, v/v)$ as the starting mobile phase, [2] S. Nagaoka, H. Ihara, J. Honbo, C. Hirayama, H. Kurisaki, S. [2] S. Nagaoka, H. Ihara, J. Honbo, C. Hirayama, H. Kurisaki, S. followed by elution of the adsorbed drug by a step
gradient to acetonitrile–water (70:30, v/v), in the
[4] I. Novák, D. Berek, CS patent 221 (1982) 197. third minute from injection. A drawback of this [5] J.H. Knox, B. Kaur, in: P.R. Brown, R.A. Hartwick (Editors), method is that many drugs are retained so strongly High-Performance Liquid Chromatography, Wiley, New that only organic solvent (pure acetonitrile, tetrahy-

drofuran or even mixtures with beyane) can elute [6] D. Berek, I. Novák, Chromatographia 30 (1990) 582. drofuran, or even mixtures with hexane) can elute $\begin{bmatrix} 6 \end{bmatrix}$ D. Berek, I. Novak, Chromatographia 30 (1990) 582.
 $\begin{bmatrix} 7 \end{bmatrix}$ O. Chiantore, I. Novák, D. Berek, Anal. Chem. 60 (1988) $\begin{bmatrix} 38 \end{bmatrix}$ corporated in the carbon pores shrinks under these [8] M. Hutta, E. Šimunicova, D. Kaniansky, J. Táčová, J. Brtko, conditions. After several tens of cycles of adsorption J. Chromatogr. 470 (1989) 223.

and elution using repeatedly aqueous and organic eluents, the back pressure of the column began to increase probably due to partial degradation of the incorporated dextran gel. Even if this limits the stability and the application area of the dextran filled carbon column, the results show that the approach might be used for preparation of novel 'restricted access' separation media.

4. Conclusions

The carbon adsorbent prepared by low temperature pyrolysis of various organic precursors is widely used as an adsorbent in gas chromatography or for bulk adsorption from liquid phase, but its use in HPLC is limited, mainly because of inhomogeneity of its surface. Therefore, almost all HPLC separations on carbonaceous adsorbents described in the literature were performed with graphitized carbon columns. This paper shows that nongraphitized carbonaceous adsorbents can be used after suitable deactivation for highly selective separations. The capillary columns packed with pyrolytic carbon combine high resolving power of the capillary device with unique selectivity of the separation medium.

Advantageous properties of the pyrolytic carbon were demonstrated on selected micro HPLC sepa-

References

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- [9] F. Belliardo, O. Chiantore, D. Berek, I. Novák, J. Chroma- [18] B.J. Fish, J. Pharm. Biomed. Anal. 11 (1993) 517. togr. 506 (1990) 371. [19] A. Wutte, G. Golbitz, S. Friebe, G.-J. Krauss, J. Chromatogr.
- [10] J.H. Knox, B. Kaur, G.R. Millward, J. Chromatogr. 352 677 (1994) 186. (1986) 3. [20] C. Hirayama, S. Nagaoka, T. Matsumo, H. Ihara, J. Honbo,
- 162. [21] A. Karlsson, C. Pettersson, J. Chromatogr. 543 (1991) 287.
- [12] B. Kaur, LC?GC Int. Mag. Sep. Sci. 3 (1990) 41. [22] K.A. Turner, LC?GC 9 (1991) 350.
- [13] E. Forgacs, T. Cserhati, Trends Anal. Chem. 14 (1995) 23. [23] A. Cappiello, P. Palma, F. Mangani, Chromatographia 32
- [14] J.H. Knox, Q.H. Wan, Ion-Exchange of Inorganic Ions and (1991) 385. Exclusion of Proteins Using Porous Graphite (PGC) Coated [24] K. Gohlin, M. Larsson, J. Microcolumn Sep. 3 (1991) 547. with Hydrophilic Polymers, 15th Int. Symp. Column LC, [25] T. Tsuda, K. Nomura, G. Nakagawa, J. Chromatogr. 248 Abstract P 137/1, Basel, 1991. (1982) 241.
-
- [16] C. Bell, E.W. Tsai, D.P. Ip, D.J. Mathre, J. Chromatogr. 675 [27] H.J. Cortes, L.W. Nicholson, J. Microcolumn Sep. 6 (1994) (1994) 248. 257.
- [17] Y. Okada, K. Koizumi, S. Kitahata, Carbohydrate Res. 254 [28] N.A. Eltekova, Chromatographia 34 (1992) 173. (1994) 1. [29] K.K. Unger, Chromatographia 31 (1991) 507.
-
-
- [11] B.J. Bassler, R.A. Hartwick, J. Chromatogr. Sci. 27 (1989) H. Kurisaki, S. Ikegami, J. Liq. Chromatogr. 18 (1995) 1509.
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- [15] M. Petro, D. Berek, I. Novák, React. Polym. 23 (1994) 173. [26] F. Belliardo, C. Lucarelli, J. Chromatogr. 535 (1990) 305.
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